

# ElectroSHOX Competent Cells

Shipping: On Dry Ice    Catalog numbers  
Batch No.: See vial    BIO-85038  $\geq 10^{10}$  cfu/ $\mu$ g of pUC19



A Meridian Life Science® Company

Store at  $-80^{\circ}\text{C}$

## Storage and stability:

ElectroSHOX Competent Cells are shipped on Dry/Blue Ice and stored at  $-80^{\circ}\text{C}$ .

## Expiry:

When stored under the recommended conditions and handled correctly, full activity of the cells is retained until the expiry date on the outer box label.

## Lot Efficiency:

This lot of electroporation competent cells was tested with an EquiBio Easyject Optima electroporator using a 0.1cm cuvette. Using settings recommended by the manufacturer and protocol as described below, actual pulse times were  $>4.5\text{ms}$  and transformation efficiencies  $>10^{10}$  cfu/ $\mu$ g pUC19 DNA.

## Product Specifications

Efficiency	Pack Size	Control Vector
$\geq 10^{10}$ cfu/ $\mu$ g of pUC19	1ml (10 x 100 $\mu$ l)	pUC19 (10pg/ $\mu$ l)

## Genotype:

F *mcrA*  $\Delta$ (*mrr-hsdRMS-mcrBC*)  $\Phi$ 80*lacZ*  $\Delta$ M15  $\Delta$ *lacX74* *recA1* *endA1* *ara*  $\Delta$ 139  $\Delta$ (*ara*, *leu*)7697 *galU* *galK*  $\lambda$ -*rpsL* (Str<sup>R</sup>) *nupG*

## Safety precautions

This product is for R&D use only, not for human use, or any other use. Please refer to the material safety data sheet for information regarding hazards and safe handling practice.

## Notes:

Research Use Only.

## Features

- Efficient transformation of large plasmids ( $>30\text{Kb}$ )
- Highest efficiency available:  $>10^{10}$  cfu/ $\mu$ g pUC19
- *recA1* and *endA1* markers to minimize recombination events and improve the quality of plasmid DNA
- Lacks *E. coli* K restriction-modification system, to facilitate cloning of methylated genomic DNA

## Applications

- Construction of cDNA and genomic DNA libraries
- Ideal for transformation of large plasmids ( $>30\text{Kb}$ )
- Blue/white color screening
- Construction of gene banks
- Efficient plasmid rescue from eukaryotic genomes

## Description

ElectroSHOX Competent Cells are highly efficient *E. coli*, ideal for the construction of cDNA or genomic libraries using electroporation. The *lacZ* mutation allows blue/white color screening and  $\alpha$ -complementation of recombinants. The *recA1* and *endA1* markers minimize recombination events and improve the quality and yield of plasmid DNA. In order to facilitate cloning of methylated genomic DNA, ElectroSHOX lacks *E. coli* K restriction-modification systems, and is ideal for the transformation of large plasmids ( $>30\text{Kb}$ ).

### Suggested Transformation Procedure for Optimal Results:

1. Pre-chill electroporation cuvettes, electroporation chamber (if applicable), and microcentrifuge tubes on ice.
2. Remove cells from  $-80^{\circ}\text{C}$  and thaw on ice.
3. Place 40-50 $\mu$ l of the competent cells into a chilled microcentrifuge tube. Add 1-5 $\mu$ l of sample DNA to cells. Thoroughly mix by gently pipetting and incubate on ice for approximately 1 minute. Note: For optimal results, sample DNA should be in sterile H<sub>2</sub>O or low ionic strength buffer such as TE. If a control is desired, repeat this step with 2 $\mu$ l of the provided Control Vector (pUC19) in a separate tube. Refreeze any unused cells and store at  $-80^{\circ}\text{C}$ .
4. Transfer cell mixture into a pre-chilled cuvette and pulse using settings recommended by manufacturer of electroporator. As a general guideline, maximum transformation efficiency is normally attained using cuvettes with a 0.1 cm gap with an applied voltage of  $\sim 1800$  (field strength of  $\sim 18$  kV/cm).
5. Immediately dilute pulsed cells to 1ml with SOC medium and transfer to a sterile culture tube.
6. Gently shake culture tube  $\sim 200\text{rpm}$  for 60 minutes at  $37^{\circ}\text{C}$ .
7. Plate by spreading 5-200 $\mu$ l of cell transformation mixture on LB agar plates containing appropriate antibiotic and incubate overnight at  $37^{\circ}\text{C}$ .

When performing the pUC19 control transformation, plate 5 $\mu$ l of the transformation mixture on a LB agar plate containing 100 $\mu$ g/ml ampicillin. To facilitate cell spreading, place a pool of SOC (100 $\mu$ l) onto surface of plate prior to addition of transformation mixture.

### Transformation Efficiency Calculation for Control Vector

$$\text{Transformation Efficiency (cfu}/\mu\text{g pUC19 DNA)} = \frac{\text{\# colonies (colony forming units)}}{\text{pg pUC19 transformed}} \times \frac{10^6 \text{ pg}}{\mu\text{g}} \times \frac{\text{Final volume } (\mu\text{l}) \text{ of transformation mix}}{\text{Volume plated } (\mu\text{l})}$$

### For example:

If 300 colonies were obtained after transforming 20pg of pUC19 and plating 5 $\mu$ l of the final 1ml transformation mixture, the calculated transformation efficiency would be:

$$300 \text{ cfu} \times 10^6 \text{ pg} \times 1000 \mu\text{l} = 3 \times 10^9 \text{ cfu}/\mu\text{g pUC19}$$

### Associated Products:

Product Name	Pack Size	Cat No
T4 DNA Ligase	500 Units	BIO-27026
Quick-Stick Ligase	50 Reactions	BIO-27027
IPTG	5g	BIO-37036
X-GAL	1g	BIO-37035

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