

RT-qPCR Extraction Control

Catalog numbers

| | | | |
|------------------|----------|---------|-----------------------------------|
| Batch : See vial | MDX028-1 | 500 Rxn | RT-qPCR Extraction Control Red |
| | MDX029-1 | 500 Rxn | RT-qPCR Extraction Control Orange |

Store at $-80\text{ }^{\circ}\text{C}$



Storage and stability:

RT-qPCR Extraction Control is shipped on dry ice. All kit components should be stored at $-80\text{ }^{\circ}\text{C}$ upon receipt. Excessive freeze/thawing is not recommended.

Expiry:

When stored under the recommended conditions and handled correctly, quality is retained until the expiry date on the outer box label.

Quality Control:

The RT-qPCR Extraction Control is extensively tested for quality and the absence of contamination.

Safety Precautions:

Please refer to the material safety data sheet for further information.

Notes:

This reagent has been manufactured under 13485 Quality Management System, and is suitable for research use only.

Features

- Easy validation of RNA extraction protocols
- Minimal interference with sample detection
- Includes a ready-to-use reaction mix for easy setup
- Suitable for use with blood, urine and sputum starting samples

Applications

- Monitoring of RNA extraction process in real-time PCR assays

The RT-qPCR Extraction Control enables users of diagnostic assays to validate both their extraction and RT-qPCR. Cells of a known concentration, containing the Internal Control RNA sequence are spiked into the sample tissue and RNA from the sample tissue and the RT-qPCR Extraction Control is simultaneously extracted.

Signal derived from the Internal Control RNA confirms the success of the extraction step and, as a known concentration of cells are added, RT-qPCR Extraction Control also monitors co-purification of PCR inhibitors that may cause biased or false amplification patterns.

Components

| Reagent | 500 Reactions |
|-----------------------|-----------------------|
| Internal Control RNA | 5 x 200 μL |
| Control Mix | 5 x 100 μL |
| 50 mM MgCl_2 | 1 x 1.2 mL |

Recommended Protocol

| Color coding | Internal Control RNA | Control Mix 560 | Control Mix 670 | 50 mM MgCl_2 |
|--------------|----------------------|-----------------|-----------------|-----------------------|
| Cap Colors | Purple | Yellow | Brown | Blue |

All steps should be carried out at room temperature unless otherwise stated. Conditions may vary depending on the assay and may need optimization.

Extraction step

1. Briefly spin down all tubes before opening.
2. Standard Protocol:
 - i) Spike 2 μL of RT-qPCR Extraction Control (REC) into each sample
 - ii) Follow the manufacturer's protocol for total RNA extraction
 - iii) Elute total RNA in a volume of 100 μL
3. Use 5 μL of the elution volume for a 20 μL PCR reaction.
For example: 2 μL REC spiked into sample, Total sample RNA extracted and eluted in 100 μL , 5 μL RNA template is used for a 20 μL reaction volume.

Note: This ratio (REC:Elution Vol:RNA template) must be maintained to avoid RNA Extraction Control failure

Post-extraction setup and analysis

The following real-time RT-PCR setup is recommended when the REC is to be used with the following:

SensiFAST™ Probe One-Step No-ROX Kit
SensiFAST Probe One-Step Lo-ROX Kit
SensiFAST Probe One-Step Hi-ROX Kit

1. Real-Time RT-PCR set up for SensiFAST Probe One-Step No-ROX Kit.

| Component | Supplied | Volume |
|---|----------|-------------------|
| 2x SensiFAST Probe One-Step No-ROX Mix* | No | 10 μL |
| Target Primer/Probe mix | No | X μL |
| Extracted RNA template | No | X μL |
| Control Mix** | Yes | 0.8 μL |
| 50 mM MgCl_2 | Yes | 1.2 μL |
| Reverse transcriptase | No | 0.2 μL |
| RiboSafe RNase inhibitor | No | 0.4 μL |
| Total Volume (for 1 reaction) | | 20 μL |

* This also applies to any commercial real-time RT-PCR mix with a standard MgCl_2 concentration of 3 mM.

** Vortex Control Mix tube before making up the master mix.

2. Recommended reverse transcription and PCR cycling

| Cycles | Temperature | Duration | Notes |
|--------|---------------------------------|-----------|---------------------------------|
| 1 | 42 $^{\circ}\text{C}$ | 10-20 min | Reverse transcription |
| 1 | 95 $^{\circ}\text{C}$ | 3 min | Activation |
| 30-40 | 95 $^{\circ}\text{C}$ | 10 s | Denaturation |
| | 60 $^{\circ}\text{C}^{\dagger}$ | 30 s-45 s | Annealing/Extension/Acquisition |

conditions.

\dagger The standard annealing temperature is 60 $^{\circ}\text{C}$, but may have to be optimized by the user, particularly if using an alternative commercial real-time RT-PCR mix

The results can be determined using the following guidelines:

| Result | Target | REC | Interpretation |
|--------|--------|-----|--|
| 1 | + | + | Target(s) and internal control RNA detected |
| 2 | - | + | Target(s) not detected, internal control RNA detected, indicates a successful extraction and real-time RT-PCR reaction |
| 3 | - | - | Invalid result: Target(s) and internal control RNA not detected, repeat test |
| 4 | + | - | Invalid result: Internal control not detected, repeat test |

Note:

a) Validation of multiplex PCR should be performed prior to high throughput processes

b) The negative control reaction should contain all components required for amplification of sample RNA, including REC

c) A negative control ensures no cross-reactivity with the user-assay and REC

Troubleshooting

| Problem | Possible Cause | Recommendation |
|--|-------------------------------------|---|
| Invalid Result or Internal Control failure | Not enough RNA template | The correct proportions are as follows: 2 µL REC per clinical sample and an elution volume of 100 µL. Check that the correct amount of extracted RNA template has been added to the reaction. |
| | Real-time RT-PCR mix not compatible | The REC system requires extra magnesium, adjust final concentration to 6 mM final. |
| Real-time RT-PCR failure* | RNA contained an RT inhibitor | Remove inhibitors, such as SDS, EDTA, formamide and pyrophosphate, by ethanol precipitation of RNA, including a 70% ethanol wash step. |
| | Reaction conditions not optimal | Increase the primer annealing step from 30 s up to 45 s. Increase the reverse transcription step from 10 min up to 20 min. |
| | RNA degraded | Analyze RNA on a denaturing gel to verify integrity. Ensure that all reagents are RNase-free. |
| Poor specificity in real-time RT-PCR | Primer dimers | Redesign primers to prevent self-annealing. Set up reactions on ice. |
| | Genomic DNA contamination | Treat RNA with DNase I and re-purify. If possible, use intron-spanning primers in real-time PCR. |
| Significant shift in Ct | Inefficient extraction | Alter extraction protocol |

* Shift in Ct or decrease in the fluorescence level (RFU) in the REC signal compared to the expected Ct or normalized fluorescence level

Technical Support:

If the troubleshooting guide does not solve the difficulty you are experiencing, please contact Technical Support with details of reaction setup, cycling conditions and relevant data.

Email: tech@meridianlifescience.com

SensiFAST is a trademark of Boline Reagents Ltd.

Associated Products

| Product | Pack size | Cat. No. |
|-------------------------------------|---------------|-----------|
| ISOLATE II RNA Mini Kit | 10 Preps | BIO-52071 |
| ISOLATE II RNA Plant Kit | 10 Preps | BIO-52076 |
| SensiFAST Probe One-Step Hi-ROX Kit | 500 reactions | BIO-77005 |
| SensiFAST Probe One-Step Lo-ROX Kit | 500 reactions | BIO-78005 |
| SensiFAST Probe One-Step No-ROX Kit | 500 reactions | BIO-76005 |

Boline Reagents Ltd
UNITED KINGDOM

Tel: +44 (0)20 8830 5300
Fax: +44 (0)20 8452 2822

Boline USA Inc.
USA

Tel: +1 800 327 6299
Fax: +1 901 382 0027

Boline GmbH
GERMANY

Tel: +49 (0)3371 60222 00
Fax: +49 (0)3371 60222 01

Boline (Aust) Pty. Ltd
AUSTRALIA

Tel: +61 (0)2 9209 4180
Fax: +61 (0)2 9209 4763

Boline France
FRANCE

Tel: +33 (0)1 42 56 04 40
Fax: +49 (0)9 70 06 62 10