α-Select Chemically Competent Cells Bacteriophage T1-Resistant

Shipping: On Dry Ice Catalog numbers

Exp. Date: See vial BIO-85029 ≥10⁸ cfu/µg of pUC19 (Silver Efficiency)

Batch No.: See vial BIO-85030 ≥10⁹ cfu/µg of pUC19 (Gold Efficiency)



Store at -80°C

Storage and stability:

 α -Select Chemically Competent Cells Bacteriophage T1-Resistant are shipped on Dry Ice and can be stored for up to 6 months at -80 °C.

Product Specifications:

 Efficiency
 Pack Size
 Control DNA

 Silver
 2ml (10 x 200µl)
 pUC19 (10pg/µl)

 Gold
 1ml (20 x 50µl)
 pUC19 (10pg/µl)

Genotype:

F⁻ deoŘ endA1 recA1 relA1 gyrA96 hsdR17(r_k⁻, m_k⁺) supE44 thi-1 phoA Δ(lacZYA-argF) U169 Φ80/acZΔM15 l⁻

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- 1. This product insert is a declaration of analysis at the time of manufacture.
- 2. Research Use Only.

Features

- Bacteriophage T1-Resistant chemically competent cells
- Two efficiencies: ≥10⁸ or ≥10⁹ cfu/µg of DNA
- Accommodate larger plasmids

Applications

- Blue/white color screening
- Construction of gene banks
- Generation of cDNA libraries using plasmid-derived vectors
- High quality plasmid preparation
- Hosting H13mp cloning vectors

Description

 α -Select Competent Cells contain a *lacZ* marker that provides α -complementation of the ß-galactosidase gene for blue/white color screening. The cells are ideal for generating cDNA libraries and subcloning. α -Select Competent Cells also provide *rec*A1 and *end*A1 markers to minimize recombination and enhance the quality of plasmid DNA. pUC19 DNA is also provided as a positive control.

Both Silver and Gold Efficiency Chemically Competent Cells are available as bacteriophage T1-Resistant strains. Many laboratories have experienced bacteriophage T1 outbreaks, as T1 attacks *E. coli* and spreads rapidly. α-Select T1-Resistant cells protect samples from bacteriophage infection.

Suggested Transformation Procedure for Optimal Results:

- 1. Remove cells from -80°C and let thaw on wet ice.
- Gently mix cells by lightly flicking tube. Aliquot ~50-100µl of cells into chilled, 17 x 100mm polypropylene tube(s), e.g., Falcon 2059. Unused cells may be refrozen, but a small drop in efficiency may result. For optimal recovery, refreeze cells in a dry ice/ ethanol bath prior to -80°C storage.
- Add DNA solution (≤5µl per 50µl cells) to cell suspension and gently swirl tube(s) for a few seconds to mix. If a control is desired, repeat this step with 2µl of the provided pUC19 in a separate tube.
- 4. Incubate on ice for 30 minutes.
- Place tube(s) in 42°C water bath for ~30 to 45 seconds without shaking. For 50µl aliquots in Falcon 2059 tubes, 30 seconds is recommended for maximum efficiency.
- 6. Replace tube(s) on ice for ~2 minutes.
- Dilute transformation reaction(s) to 1ml by addition of 900-950µl SOC. SOC Medium: 2% Tryptone, 0.5% Yeast Extract, 0.4% glucose, 10mM NaCl, 2.5mM KCl, 10mM MgCl₂ & 10mM MgSO₄.
- 8. Shake tube(s) ~200 rpm for 60 minutes at 37°C.
- Plate by spreading 5-200µl of cell transformation mixture on LB agar plates containing appropriate antibiotic and incubate overnight at 37°C

When performing the pUC19 control transformation, plate 5μ I of the transformation mixture on a LB agar plate containing 100μ g/mI ampicillin. To facilitate cell spreading, place a pool of SOC (100μ I) onto surface of plate prior to addition of transformation mixture.

Transformation Efficiency Calculation for Control DNA

Transformation Efficiency (cfu/µg pUC19 DNA) # colonies

(colony forming units)

pg pUC19
transformed

Χ 10⁶ pg μg

Final volume (µl) of transformation mix Volume plated (µl)

For example:

If 40 colonies were obtained after transforming 20pg of pUC19 and plating 5µl of the final 1ml transformation mixture, the calculated transformation efficiency would be:

40cfu 20pg pUC19 X 10⁶ pg

X 1000μl

= 4 x 10⁸ cfu/µg pUC19

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Associated Products:

Product Name	Pack Size	Cat No
T4 DNA Ligase	500 Units	BIO-27026
Quick-Stick Ligase	50 Reactions	BIO-27027
IPTG	5g	BIO-37036
X-GAL	1g	BIO-37035

Product Citations:

- 1. Stat, M., et al. PLoS One 6(1), e15854 (2011)
- 2. Hornsey, M. et al. J. Antimicro. Agents 35(5), 478-81 (2010)
- Almeida-Vega, S., et al. Am. J. Physiol. Gastrointest. Liver Physiol, 296, 414-23 (2009)
- 4. Schultz, J. K., et al. J. Heredity 100(1), 25-33 (2009)
- 5. Catlow, K., et al. JBC 282, 17069-77 (2007)
- Donato, J.J., et al. PLoS Genet 2(9), e141 (2006)

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