ACCUZYME™ Mix

Shipping: On Dry/Blue Ice Catalog numbers

Batch No.: See vial BIO-25028: 500 x 50μl reactions: (10 x 1.25ml)

Concentration: 2x

BIOLINE Meridian Life Science® Company Store at -20°C

The ACCUZYME Mix is shipped on dry/blue ice. On arrival store at -20°C for optimum stability. Repeated freeze/thaw cycles should be avoided.

Expirv:

When stored under the recommended conditions and handled correctly, full activity of the kit is retained until the expiry date on the outer box label.

Safety precautions:

Storage and stability:

Please refer to the material safety data sheet for further information.

Quality control specifications:

ACCUZYME Mix and its components are extensively tested for activity, processivity, efficiency, sensitivity, absence of nuclease contamination and absence of nucleic acid contamination prior to release.

Notes:

Research use only.

ACCUZYME is a trademark of Bioline Reagents Ltd.

Features

- High fidelity coupled with high yield
- Amplifies fragments up to 5Kb
- Convenient pre-mixed, pre-optimized 2x solution
- Reduced risk of contamination
- Dramatically decreases the time required for reaction set-up
- Reproducible results

Applications

- Ideal for ultra-high fidelity for subsequent cloning
- Blunt-end cloning
- Site-directed mutagenesis

Description

ACCUZYME™ Mix is a convenient ready-to-go 2x reaction mix designed to maximize experiment reproducibility. ACCUZYME Mix contains ACCUZYME DNA Polymerase, MgCl₂ and ultra-pure dNTPs manufactured by Bioline. The mix is optimized and ready-to-use, the user is simply required to add water, template and primers.

ACCUZYME Mix dramatically reduces the time needed to set up reactions, thereby minimizing the risk of contamination. Greater reproducibility is ensured, by a reduction in the number of pipetting steps that can lead to pipetting errors. ACCUZYME Mix is supplied with an additional 50mM MgCl₂ solution for optional optimization of reaction conditions.

Components

	500 Reactions
ACCUZYME™ Mix, 2x	10 x 1.25ml
50mM MgCl ₂ Solution	1.2ml

Standard ACCUZYME Mix Protocol

The following protocol is for a standard 50µl reaction and can be used as a starting point for reaction optimization. Please refer to the Important Considerations and PCR Optimization section.

PCR reaction set-up:

ACCUZYME™ Mix, 2x	25µI
Primers 20mM each	1μl
Template	as required
Water (ddH ₂ O)	up to 50µl

PCR cycling conditions

Step	Temp.	Time	Cycles	
Initial denaturation	95-98°C	3 min	1	
Denaturation	95-98°C	15s		
Annealing*	55-60°C	15s 25-35		
Extension	72°C	1.5 - 2 min/kb		

^{*}Annealing temperature is primer dependent

The conditions above are intended for use as a guide only: conditions will vary from reaction to reaction and may need optimization.

Important considerations and PCR optimization

The optimal conditions will vary from reaction to reaction and are dependent on the template/primers used.

 Mg^{2+} concentration: The Mg^{2+} concentration in the 2x mix is 4mM (2mM final concentration), this is the optimum concentration for ACCUZYME Mix for most PCR reactions and should only be adjusted if necessary.

Primers: Forward and reverse primers are generally used at the final concentration of 0.2-0.6µM each. As a starting point, we recommend using 0.4µM final concentration (i.e. 20pmol of each primer per 50µl reaction volume). Too high a primer concentration can reduce the specificity of priming, resulting in non-specific products.

When designing primers we recommend using primer-design software such as Primer3 (http://frodo.wi.mit.edu/primer3) or visual OMPTM (http://dnasoftware.com). Primers should have a melting temperature (Tm) of approximately 60°C.

Template: The amount of template in the reaction depends mainly on the type of DNA used. For templates with low structural complexity, such as plasmid DNA, we recommend using 50pg-10ng DNA per 50µl reaction volume. For eukaryotic genomic DNA, we recommend a starting amount of 200ng DNA per 50µl reaction, this can be varied between 5ng-500ng. It is important to avoid using template re-suspended in EDTA-containing solutions (e.g. TE buffer) since EDTA chelates free Mg²⁺.

Troubleshooting Guide

Problem	Possible Cause	Recommendation	
	Missing component	- Check reaction set-up and volumes used	
	Defective component	- Check the aspect and the concentrations of all components as well as the storage conditions. If necessary test each component individually in controlled reactions	
No PCR	Cycling conditions not optimal	 Decrease the annealing temperature Run a temperature gradient to determine the optimal annealing temperature Increase the extension time, especially if amplifying a long target Increase the number of cycles 	
	Difficult template e.g. GC or ATrich, or high level of secondary structure	 Increase initial denaturation time to 5 minutes Increase denaturation time 	
	Excessive cycling	- Decrease the number of cycles	
Smearing	Extension time too long	nsion time too long - Decrease the extension time	
or	Annealing temperature too low	- Increase the annealing temperature	
Non-Specific	Primer concentration too high	- Decrease primer concentration	
products	Contamination	Replace each component in order to find the possible source of contamination Set-up the PCR reaction and analyze the PCR product in separated areas	

Product Citations

ACCUZYME DNA Polymerase

- 1. Kitazono, A.A., *Gene* doi:10.1016/j.gene.2011.06.006 (2011).
- 2. Batchelor, D.J., et al. Am. J. Physiol. 300, R67-R75 (2011).
- 3. Chiang, C., et al. J. Bacteriol. 193, 52-62 (2011).
- 4. Chin, G.L., et al. Appl. Envir. Microbiol. 77, 3451-3460 (2011).
- Cheng, C., et al. Mol. Cell. Biol. 31, 983-997 (2011).
- 6. Chakrabarti, M., et al. Virol. J. 7, 181 (2010).
- 7. Silvestrini, F., et al. Mol.Cell. Prot., 9, 1437-48 (2010).
- 8. Williamson, D. S., et al. Appl. Microbiol. Biotechnol. 88, 143-153 (2010).
- 9. Johnson M., et al. NAR 37(14), e98 (2009).
- 10. Pacheco, A., et al. Microbiol. 155, 2021-2028 (2009).
- 11. Wilson, A. C., et al. J. Bacteriol. 190(15), 5522-5525 (2008).

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- 1. Padmashali R.M. & Andreadis. S.T., Biomaterials 32 (12), 3330-3339 (2011).
- 2. Potula, S. K., et al. Transgen. Res. 17(1), 19-32 (2008).
- 3. Jury, F., et al. Med. Microbiol. 55, 1053-1060 (2006).

Technical Support

If the troubleshooting guide does not solve the difficulty you are experiencing, please contact your local distributor or our Technical Support with details of reaction setup, cycling conditions and relevant data.

Email: tech@bioline.com

Associated Products

Product Name	Pack Size	Cat. No.
dNTP Set	4 x 25μmol	BIO-39025
dNTP Mix	500µl	BIO-39028
ACCUZYME™ DNA Polymerase	250 units (100µl)	BIO-25021

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